

Tadashi Terada · Tetsuo Ohta · Yukisato Kitamura
Keigo Ashida · Yoshiko Matsunaga · Masako Kato

Endocrine cells in intraductal papillary-mucinous neoplasms of the pancreas

A histochemical and immunohistochemical study

Received: 3 December 1996 / Accepted: 25 February 1997

Abstract The endocrine cells in intraductal papillary-mucinous neoplasms (IPN) of the pancreas have rarely been investigated. In the normal pancreatic ducts of normal pancreases ($n=5$) there were a few endocrine cells: argyrophil in 5 (100%), chromogranin A in (100%), pancreatic polypeptide (PP) in 3 (60%), and insulin in 7 (20%). These endocrine cells were scattered, and located in the basal portions of pancreatic ducts. In IPN of the pancreas ($n=9$), there were many endocrine cells: argyrophil in 7 (78%), argentaffin in 8 (89%), chromogranin A in 8 (89%), PP in 7 (78%), serotonin in 7 (78%), insulin in 4 (44%), and gastrin in 5 (56%). In invasive ductal adenocarcinoma of the pancreas ($n=6$), many endocrine cells were also detected: argyrophil cells in (67%), chromogranin A in 3 (50%), insulin in 3 (50%), glucagon in 4 (67%), and somatostatin in 3 (50%). In positive cases, endocrine cells were situated under or among the neoplastic cells and the proportion of endocrine cells in IPN was less than 5% of the total neoplastic cell population. These data show that normal pancreatic ducts contain endocrine cells and that IPN frequently contain argyrophil, argentaffin, chromogranin A, and hormone-containing endocrine cells. These data also suggest that endocrine differentiation occurs during neoplastic transformation and progression of IPN of the pancreas.

Introduction

As neuron and endocrine cells have been found to have many common features, the term “neuroendocrine” has been applied to them [6], though they have been found in many organs other than the endocrine glands. They have

common histochemical features, including amino uptake, serotonin positivity, and affinity to potassium dichromate (chromaffin) [6], and characteristically have an affinity to silver. Electron microscopy and immunohistochemistry have revealed that these cells have characteristic secretory granules and also contain various peptide hormones [6]. Endocrine cells are present among the normal cells of non-endocrine organs as well as in cancerous non-endocrine organs [3]. These cells secrete several hormones that regulate cell growth and differentiation via autocrine and/or paracrine loops [5, 21, 28]. The receptors for these hormones are also present in cells of the non-endocrine organs [14, 34]. In the exocrine pancreas, endocrine cells have been reported to be present in the pancreatic ducts of the normal pancreas and pancreases affected by chronic pancreatitis [2, 4, 30] as well as in pancreatic ductal carcinomas [2, 4, 7, 9, 11, 22, 30].

Intraductal papillary-mucinous neoplasm (IPN) of the pancreas is a rare and unique form of pancreatic neoplasm. It is characterized by intraductal papillary epithelial proliferations with dilatation of the main pancreatic duct and/or its major branches [1, 8, 13, 16–19, 23–27, 29, 31–33, 36]. A large amount of mucin usually fills the dilated pancreatic ducts. IPN usually shows a benign clinical course, but malignant transformation has been reported [1, 8, 13, 16–19, 23–27, 29, 31–33, 36]. Histopathologically, IPN shows variable histology, ranging from tall, highly differentiated, columnar mucin-containing cells to carcinoma in situ [1, 8, 13, 16–19, 23–27, 29, 31–33, 36], which reflects its variable biological behaviour. In the recent revised WHO classification [10], IPN was classified as benign, borderline and malignant. Much effort has been devoted to characterization of this neoplasm. It has been shown that IPN occasionally expresses oncofetal antigens [17, 18, 26, 29, 31, 32] and has altered MUC apomucin expression [33]. In addition, abnormalities of oncogenes and anti-oncogenes have been reported in IPN [13, 26, 29, 32]. However, endocrine cells have rarely been reported in IPN [16–19].

In this study, therefore, we investigated the distribution of argyrophil and argentaffin cells by histochemistry

T. Terada (✉) · Y. Kitamura · K. Ashida · Y. Matsunaga · M. Kato
Second Department of Pathology,
Tottori University Faculty of Medicine,
Nishimachi 86, Yonago 683, Japan
Fax (81) 859-34-8348

T. Ohta
Second Department of Surgery,
Kanazawa University School of Medicine, Kanazawa, Japan

We collected nine cases of IPN of the pancreas from autopsy (three) and surgical (six) cases seen at our laboratories and affiliated hospitals during 1986–1995 (Table 1). According to the revised WHO classification [10], cases 1–4 in Table 1 were classified as intraductal papillary-mucinous adenoma, cases 5–7, as intraductal papillary-mucinous adenoma with moderate atypia, case 8, as intraductal papillary-mucinous carcinoma (non-invasive), and case 9, as invasive intraductal papillary-mucinous carcinoma.

We also obtained five normal pancreases from recent autopsy files at our laboratories and six invasive ductal adenocarcinomas of the pancreas from recent surgical files. Many tissue specimens were obtained from each pancreatic specimen; they were fixed in 10% formalin and embedded in paraffin. Several 3- μ m sections were obtained from each paraffin block. One of them was stained with haematoxylin and eosin, and two with Grimelius' technique for argyrophil cells and with Masson-Fontana's technique for argentaffin cells. The rest were subjected to immunohistochemical study for chromogranin A and several gut hormones.

Chromogranin A and several gut hormones (Table 2) were stained immunohistochemically by the standard avidin-biotin-peroxidase complex (ABC) method. In brief, after deparaffinization, endogenous peroxidase activity was abolished in 100% methanol containing 0.3% H_2O_2 . The sections were then treated at 4°C overnight with antibody solution as described in Table 2. The anti-chromogranin A reacts with a 68-kDa protein that is associated with endocrine secretory granules [35]. The sections were then treated with secondary biotinylated antibodies (Vector Lab, Burlingame, Calif.) for 2 h, followed by treatment with the ABC (Vectastain ABC Elite Kit, Vector Lab) for 1 h. Reaction products were developed by immersing the sections in 3,3'-diaminobenzidine tetrahydrochloride solution containing 0.03% H_2O_2 . Nuclei were lightly counterstained with haematoxylin. Reactions products were not present when nonimmune serum or phosphate-buffered saline was used instead of the primary antibodies.

Endocrine cells positive with Grimelius' and Masson-Fontana' stains were termed argyrophil and argentaffin cells, respectively. Endocrine cells positive for chromogranin A, a 68-kDa protein associated with endocrine secretory granules [35], were termed as chromogranin A cells. Endocrine cells positive for individual gut hormones were collectively termed hormone-containing cells, and they were subclassified as pancreatic polypeptide (PP), serotonin, insulin, gastrin, glucagon, somatostatin, and vasoactive intestinal polypeptide (VIP) cells.

The results are summarized in Table 1.

Argyrophil cells were scattered in the pancreatic ducts in all normal pancreases (Fig. 1A), but no argentaffin cells were found in any of these cases. Chromogranin A cells were scattered in the pancreatic ducts in all cases (Fig. 1B). PP and insulin cells were also scattered in three (60%) and one (20%) of the normal cases, respectively (Table 1). However, there were no serotonin, gastrin, glucagon, VIP, and somatostatin cells. The argyrophil, chromogranin A, PP or insulin cells accounted for

Table 1 Endocrine cells in normal pancreatic ducts, intraductal papillary-mucinous neoplasm of the pancreas, and pancreatic invasive ductal adenocarcinoma (*M*, male, – negative, +positive, *PP* pancreatic polypeptide, *VIP* vasoactive intestinal polypeptide; Figures in parentheses before sex show age in years)

| Endocrine cells | Normal pancreas (<i>n</i> =5) | Intraductal papillary-mucinous neoplasm | | | | | | | | | Total | Invasive ductal adenocarcinoma (<i>n</i> =6) |
|-----------------|-----------------------------------|---|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|-----------|---|
| | | Case 1 (71, M) | Case 2 (62, M) | Case 3 (91, M) | Case 4 (59, M) | Case 5 (69, M) | Case 6 (62, M) | Case 7 (72, M) | Case 8 (72, M) | Case 9 (81, M) | | |
| Argyrophil | 5/5 (100%) | + | + | — | + | — | + | + | + | + | 7/9 (78%) | 4/6 (67%) |
| Argentaffin | 0/5 (0%) | + | + | — | + | + | + | + | + | + | 8/9 (89%) | 0/6 (0%) |
| Chromogranin A | 5/5 (100%) | + | + | — | + | + | + | + | + | + | 8/9 (89%) | 3/6 (50%) |
| PP | 3/5 (60%) | — | + | + | + | — | + | + | + | + | 7/9 (78%) | 0/6 (0%) |
| Serotonin | 0/5 (0%) | + | + | — | — | + | + | + | + | + | 7/9 (78%) | 0/6 (6%) |
| Insulin | 1/5 (20%) | — | + | + | — | + | + | + | — | — | 4/9 (44%) | 3/6 (50%) |
| Gastrin | 0/5 (0%) | — | + | — | + | — | + | + | — | + | 5/9 (56%) | 0/6 (0%) |
| Glucagon | 0/5 (0%) | — | — | — | — | — | — | — | — | — | 0/9 (0%) | 4/6 (67%) |
| Somatostatin | 0/5 (0%) | — | — | — | — | — | — | — | — | — | 0/9 (0%) | 0/6 (0%) |
| VIP | 0/5 (0%) | — | — | — | — | — | — | — | — | — | 0/9 (0%) | 3/6 (50%) |

Table 2 Primary antibodies used in the present study (M/P monoclonal/polyclonal, PP pancreatic polypeptide, VIP vasoactive intestinal polypeptide)

| Antibodies | M/P | Source | Dilution |
|-------------------------|-----|-------------------------------|----------|
| Chromogranin A (LK2H10) | M | Immunotech, Marseille, France | ×1 |
| PP | P | BioGenex, Dublin, Ireland | ×1 |
| Serotonin | M | Dakopatts, Glostrup, Denmark | ×50 |
| Insulin | P | Dakopatts, Glostrup, Denmark | ×200 |
| Gastrin | P | Dakopatts, Glostrup, Denmark | ×250 |
| Glucagon | P | Dakopatts, Glostrup, Denmark | ×150 |
| Somatostatin | P | Dakopatts, Glostrup, Denmark | ×300 |
| VIP | P | BioGenex, Dublin, Ireland | ×1 |

less than 3% of the total pancreatic duct cell population. The density of the endocrine cells was higher in large pancreatic ducts than in small pancreatic ducts and ductules.

In the nine IPNs of the pancreas, the main pancreatic duct and/or its major branches were severely dilated with proliferations of the duct cells (Fig. 2). Argyrophil (Fig. 3A), argentaffin (Fig. 3B) and chromogranin A cells (Fig. 3C) were present in seven (78%), eight (89%) and eight (89%) of the IPNs, respectively (Table 2). There were many hormone-containing cells; PP cells in seven cases, or 78% (Fig. 3D), serotonin cells in seven, or 78% (Fig. 3E), insulin cells in four, or 44% (Fig. 3F), and gastrin cells in five, or 56% (Fig. 3G). There were no glucagon, VIP or somatostatin cells. In positive cases, these endocrine cells were situated under or among the neoplastic cells, and the endocrine cells accounted for less than 5% of the total neoplastic cell population.

In invasive ductal adenocarcinoma of the pancreas, argyrophil (Fig. 4A), argentaffin and chromogranin A cells (Fig. 4B) were present in four (67%), none (0%) and three (50%) of the six cases, respectively (Table 1). There were also hormone-containing cells; insulin cells in three (50%), glucagon cells in four (67%) and somatostatin cells in three (50%). There was no immunoreactivity of PP, serotonin, gastrin or VIP in cancer cells in any of these cases. In positive cases, the argyrophil, chromogranin and hormone-containing cells were located under or among the carcinoma cells, and these endocrine cells made up a proportion ranging approximately from 5% to 50% of the total carcinoma cell population. In the cancerous stroma there were endocrine cells that were considered to the residual islet cells.

Discussion

Endocrine cells have been recognized by their affinity to silver (argyrophil and argentaffin) and by immunohistochemical demonstration of chromogranin A, a 68-kDa protein associated with secretory granules [35]. Recent advances in immunohistochemistry have made it possible to reveal the hormones that are produced in the cells of the gut [6]. We examined argyrophil, argentaffin and

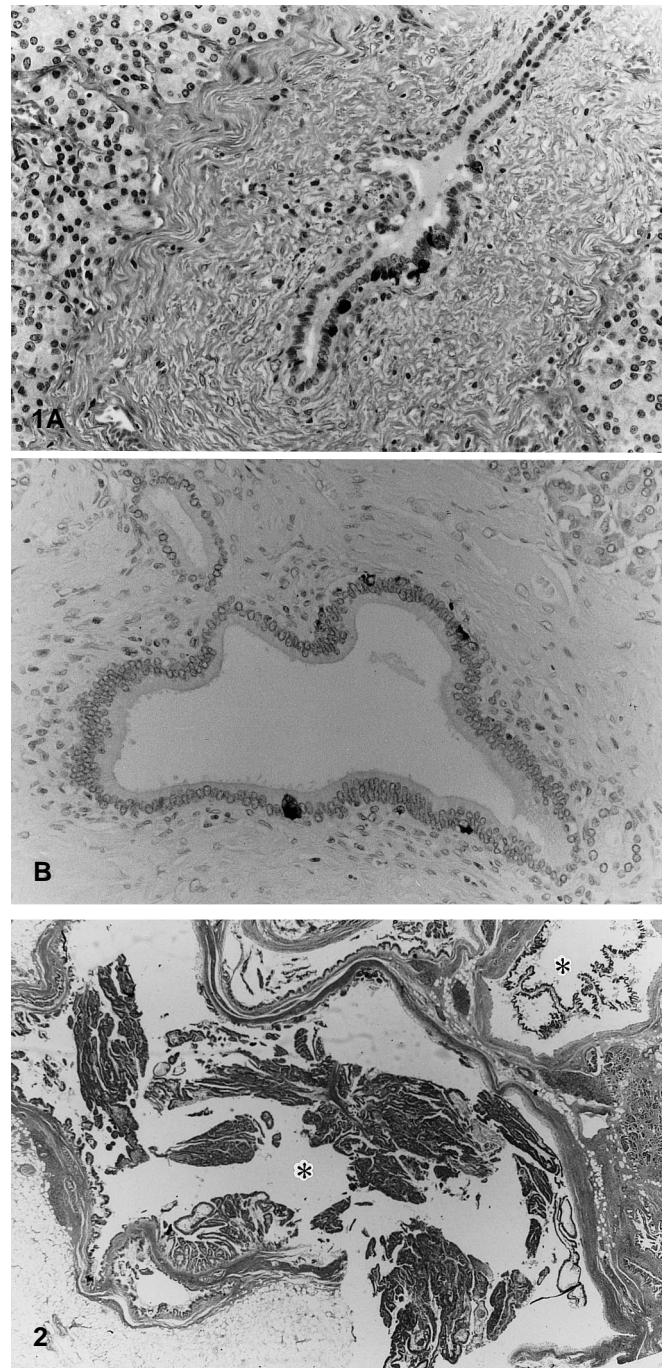


Fig. 1A, B Endocrine cells in the normal pancreatic duct of the normal pancreas. **A** Argyrophil and **B** chromogranin A cells are scattered in the basal portion of the epithelium of the normal pancreatic duct. **A** Grimelius stain, ×200. **B** Immunostaining for chromogranin A, ×200

Fig. 2 Low-power microscopic features of an intraductal papillary-mucinous neoplasm of the pancreas. The main pancreatic ducts (asterisks) are grossly dilated, and there are polyp-like protrusions within the pancreatic ducts. Haematoxylin and eosin, ×20

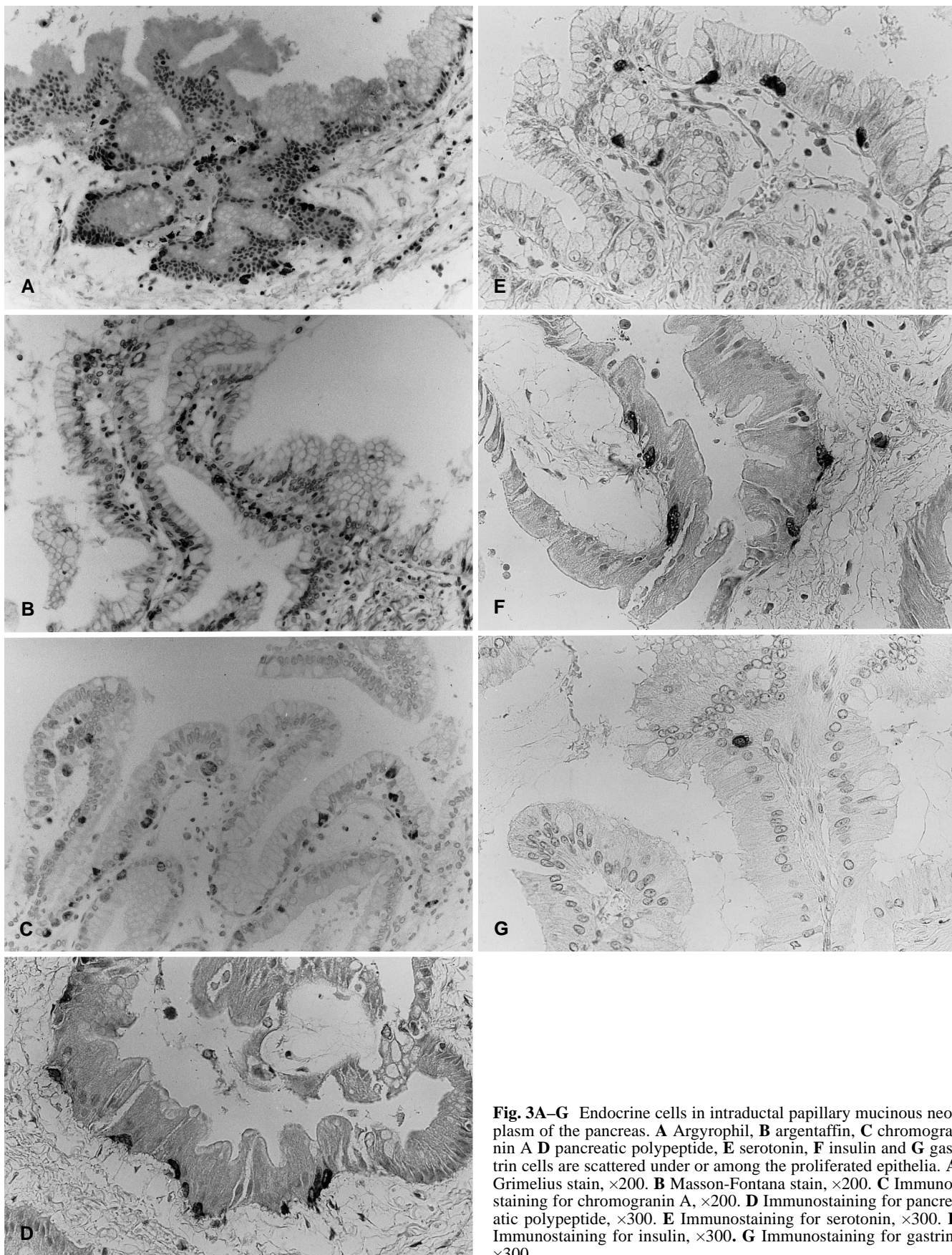


Fig. 3A–G Endocrine cells in intraductal papillary mucinous neoplasm of the pancreas. **A** Argyrophil, **B** argentaffin, **C** chromogranin **A** **D** pancreatic polypeptide, **E** serotonin, **F** insulin and **G** gastrin cells are scattered under or among the proliferated epithelia. **A** Grimelius stain, $\times 200$. **B** Masson-Fontana stain, $\times 200$. **C** Immunostaining for chromogranin A, $\times 200$. **D** Immunostaining for pancreatic polypeptide, $\times 300$. **E** Immunostaining for serotonin, $\times 300$. **F** Immunostaining for insulin, $\times 300$. **G** Immunostaining for gastrin, $\times 300$

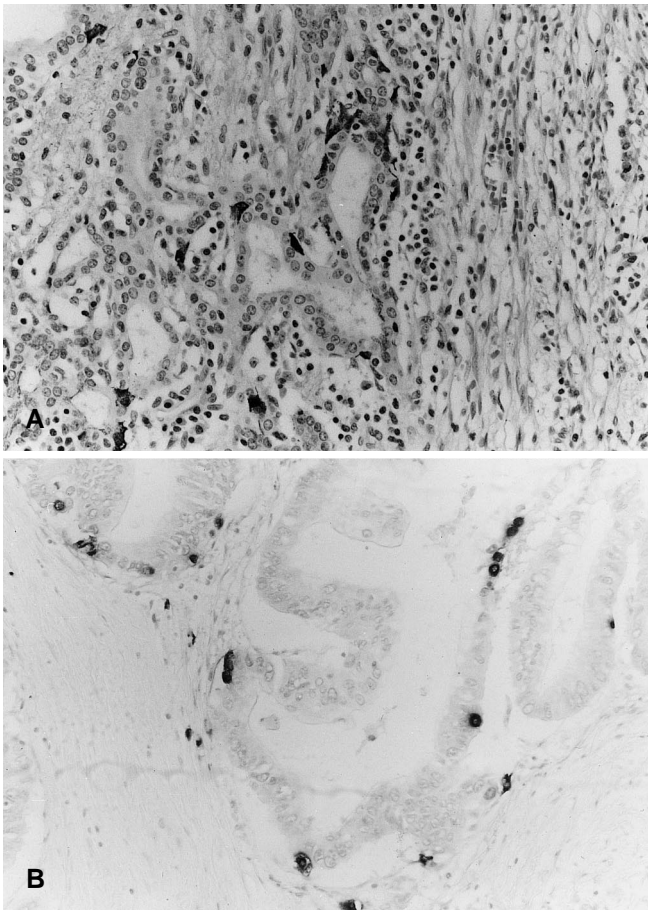


Fig. 4A, B Endocrine cells in invasive ductal adenocarcinoma of the pancreas. **A** Argyrophil and **B** chromogranin A cells are scattered under or among the carcinoma cells. **A** Grimelius stain, $\times 200$. **B** Immunostaining for chromogranin A, $\times 200$

chromogranin A cells, and also gut hormones. A high prevalence of endocrine cells was demonstrated by argyrophil staining and immunohistochemical staining for chromogranin A, PP, and serotonin, suggesting that these stains are useful for the detection of endocrine cells in the pancreatic ducts and their lesions.

In this study, the percentage of chromogranin A-positive cells was very similar to that of argyrophil cells in normal pancreas, IPN and invasive ductal adenocarcinoma. In this regard, Rindi et al. [24], using histochemistry, immunohistochemistry and dot-blot techniques, demonstrated that argyrophil cells reacted with chromogranin A antibody and that there was a close relationship between chromogranin A and argyrophil cells. This result was confirmed by Lundqvist et al. [15], who demonstrated using histochemistry and dot-blot studies that chromogranin A caused an argyrophil but not an argentaffin reaction. These observations may indicate that argyrophil and chromogranin A cells are the same, which might explain the similar incidence of argyrophil cells and chromogranin A cells in the present study.

Our study revealed that argyrophil, chromogranin, PP and insulin cells were scattered in normal pancreatic

ducts. These findings are compatible with previous studies [2, 4, 30] and indicate that endocrine cells are present in the pancreatic duct system under normal conditions. The presence of these cells in normal ducts may contribute to the maintenance of the microenvironment of pancreatic ducts under normal conditions. We found no argentaffin, serotonin, gastrin, glucagon, VIP or somatostatin cells in the normal pancreatic ducts. In general, endocrine cells and their neoplasms (carcinoid tumour) of foregut organs are argyrophil but not argentaffin [12]. This may explain why argentaffin cells were not present in normal pancreatic ducts. The reason for the absence of other hormones is not clear.

The present study showed that argyrophil, argentaffin and chromogranin A cells were present in many cases of IPN. In addition, there were many hormone-containing cells in IPN. There have been few reports on the endocrine cells of IPN, and there have been no systematic studies on endocrine cells in IPN. Milchgrub et al. [16] mentioned that argyrophil and serotonin cells were present in one of four cases of IPN, Nishihara et al. [19] noted the presence of endocrine cells in one of two IPNs, and Nagai et al. [18] reported the presence of serotonin and chromogranin A cells. Our data show that most IPNs of the pancreas harbour endocrine cells. The origin of these cells is obscure; they may be pre-existing endocrine cells or, alternatively, they may be the result of endocrine differentiation of IPN. The latter explanation seems more likely, as no argentaffin, serotonin or gastrin cells were present in normal pancreatic ducts, while such cells were found in IPN. Although the function of endocrine cells in IPN is not known, it seems possible that hormones secreted from the endocrine cells may regulate tumour cell growth via an autocrine and/or paracrine loop, as is suspected in other tumours [5, 21, 28]. However, the role of endocrine cells in IPN may be limited as they occupied less than 5% of the total neoplastic cell population.

Argyrophil and chromogranin A cells were frequently present in cases of invasive ductal adenocarcinoma of the pancreas. In addition, there were many hormone-containing cells in cases of invasive ductal adenocarcinoma. The presence of these endocrine cells has already been described [2, 4, 7, 9, 11, 22, 30]. In our series, there was not even one case of a ductal adenocarcinoma that contained argentaffin cells, suggesting that transdifferentiation to argentaffin cells does not occur in ductal adenocarcinoma of the pancreas. The endocrine cells in ductal adenocarcinoma might arise through endocrine differentiation of carcinoma cells, as previously reported in various malignant neoplasms [3, 20]. Alternatively, however, these endocrine cells may be non-neoplastic; in several cancers endocrine cells have no proliferative activity [20] and are usually absent from metastases. The hormones secreted from endocrine cells may influence carcinoma cell growth in pancreatic ductal adenocarcinoma.

References

1. Adsay NV, Adair CF, Heffess CS, Klimstra DS (1996) Intraductal oncocystic papillary neoplasm of the pancreas. *Am J Surg Pathol* 20:980–994
2. Bommer G, Friedle U, Heitz PU, Klöppel G (1980) Pancreatic PP cell distribution and hyperplasia: immunohistochemical morphology in the normal pancreas, in chronic pancreatitis and pancreatic carcinoma. *Virchows Arch [A]* 387:319–331
3. Bosman FT (1989) Endocrine cells in non-endocrine tumours. *J Pathol (Lond)* 159:181–182
4. Chen J, Baithun SI, Pollack DJ, Berry CL (1988) Argyrophilic and hormone immunoreactive cells in normal and hyperplastic pancreatic ducts and exocrine pancreatic carcinoma. *Virchows Arch [A]* 413:399–405
5. Cuttitta F, Carney DN, Mushline J (1985) Bombesin-like peptides can function as autocrine growth factor in human small-cell lung cancer. *Nature* 316:832–836
6. Delellis RA, Dayal Y (1992) Neuroendocrine system. In: Sternberg SS (ed) *Histology for pathologists*. Raven Press, New York, pp 347–362
7. Eusebi V, Cappella C, Bondi A, Sessa F, Vezzadini P, Mancini AM (1981) Endocrine cells in pancreatic exocrine carcinoma. *Histopathology* 5:599–613
8. Furukawa T, Takahashi T, Kobari M, Matsuno S (1992) The mucus-hypersecreting tumor of the pancreas. *Cancer* 70:1505–1513
9. Kay D, Delellis RA, Dayal Y, Lloyd RV, Duggan MA, Tallberg K, Sternberg SS, Wolfe HJ (1985) Ductal adenocarcinoma of the pancreas with neuroendocrine cells: an immunohistochemical study. *Lab Invest* 52:33A–34A
10. Klöppel G, Solcia E, Longnecker DS, Cappella C, Sobin LH (1996) *Histology typing of tumours of the exocrine pancreas*. (World Health Organization Series) Springer, New York Berlin Heidelberg
11. Kodama T, Mori W (1983) Morphological behavior of carcinoma of the pancreas. 2. Argyrophil cells and Langerhans' islets in the carcinomatous tissues. *Acta Pathol Jpn* 33:483–493
12. Lechago J (1978) Endocrine cells of the gastrointestinal tract and their pathology. *Pathol Annu* 2:329–350
13. Lemoine N, Jain S, Hughes CM, Staddon SL, Maillet B, Hall PA, Klöppel G (1992) *Ki-ras* oncogene activation in preinvasive pancreatic cancer. *Gastroenterology* 102:230–236
14. Longnecker DS (1991) Hormones and pancreatic cancer. *Int J Pancreatol* 9:81–86
15. Lundqvist M, Arnberg H, Candell J, Malmgren M, Wilander E, Grimelius L, Oberg K (1990) Silver stains for identification of neuroendocrine cells: a study of the chemical background. *Histochem J* 22:615–623
16. Milchgrub S, Campuzano M, Casillas J, Albores-Saavedra J (1992) Intraductal carcinoma of the pancreas. *Cancer* 69:651–656
17. Morohoshi T, Kanda M, Asanuma K, Klöppel G (1989) Intraductal papillary neoplasms of the pancreas. *Cancer* 64:1329–1335
18. Nagai E, Ueki T, Chijiwa K, Tanaka M, Tsuneyoshi M (1995) Intraductal papillary mucinous neoplasms of the pancreas associated with so-called "mucinous ductal ectasia": histochemical and immunohistochemical analysis of 29 cases. *Am J Surg Pathol* 19:576–589
19. Nishihara K, Fukuda T, Tsuneyoshi M, Kominami T, Maeda S, Saku M (1993) Intraductal papillary neoplasm of the pancreas. *Cancer* 72:689–696
20. Ooi A, Hayashi H, Katsuda S, Nakanishi I (1992) Gastric carcinoma cells with endocrine differentiation show no evidence of proliferation. *Hum Pathol* 23:736–741
21. Palmer Smith J, Solomon TE, Bagheri S (1990) Cholecystokinin stimulates growth of human pancreatic adenocarcinoma. *Dig Dis Sci* 35:1337–1384
22. Pour PM, Permert J, Mogaki M, Fujii H, Kazakoff K (1993) Endocrine aspects of exocrine cancer of the pancreas: their patterns and suggested biological significance. *Am J Clin Pathol* 100:223–230
23. Rickaert F, Cremer M, Deviere J, Tavares L, Lambiliotte JP, Schröder S, Wurbs D, Klöppel G (1991) Intraductal mucin-hypersecreting neoplasms of the pancreas: a clinicopathologic study of eight patients. *Gastroenterology* 101:512–519
24. Rindi G, Buffa R, Sessa F, Tortora O, Solcia E (1986) Chromogranin A, B and C immunoreactivities of mammalian endocrine cells: distribution, distinction from costored hormones/prohormones and relationship with the argyrophil component of secretory granules. *Histochemistry* 85:19–28
25. Santini D, Campione O, Salerno A, Gullo L, Mazzoleni G, Leone O, Martinelli G, Marrano D (1995) Intraductal papillary-mucinous neoplasm of the pancreas: a clinicopathologic entity. *Arch Pathol Lab Med* 119:209–213
26. Satoh K, Sasano H, Shimesegawa T, Koizumi M, Yamazaki T, Mochizuki F, Kobayashi N, Okano T, Toyota T, Sawai T (1993) An immunohistochemical study of the *c-erbB-2* oncogene product in intraductal mucin-hypersecreting neoplasms and in ductal cell carcinomas of the pancreas. *Cancer* 72:51–56
27. Satoh K, Ohtani H, Shimesegawa T, Koizumi M, Sawai T, Toyota T (1994) Infrequent stromal expression of gelatinase A and intact basement membrane in intraductal neoplasms of the pancreas. *Gastroenterology* 107:1488–1495
28. Scholar EM, Paul S (1991) Stimulation of tumor cell growth by vasoactive intestinal peptide. *Cancer* 67:1561–1564
29. Sessa F, Solcia E, Capella C, Bonato M, Scarpa A, Zamboni G, Pellegata NS, Ranzani GN, Rickaert F, Klöppel G (1994) Intraductal papillary-mucinous tumours represent a distinct group of pancreatic neoplasm: an investigation of tumour cell differentiation and *K-ras*, *p53* and *c-erbB-2* abnormalities in 26 patients. *Virchows Arch* 425:357–367
30. Suda K, Hashimoto K (1979) Argyrophil cells in the exocrine pancreas. *Acta Pathol Jpn* 29:413–419
31. Terada T, Nakanuma Y (1996) Expression of mucin carbohydrate antigens (T, Tn and sialyl-Tn) and MUC 1 gene product in intraductal papillary mucinous neoplasm of the pancreas. *Am J Clin Pathol* 105:613–620
32. Terada T, Ohta T, Nakanuma Y (1996) Expression of oncogene products, anti-oncogene products and oncofetal antigens in intraductal papillary-mucinous neoplasm of the pancreas. *Histopathology* 29:355–361
33. Terada T, Ohta T, Sasaki M, Nakanuma Y, Kim YS (1996) Expression of MUC apomucins in normal pancreas and pancreatic tumours. *J Pathol (Lond)* 180:160–165
34. Upp JRJ, Singh P, Townsent CMJ (1989) Clinical significance of gastrin receptors on human colon cancers. *Cancer Res* 49:488–492
35. Wilson BS, Lloyd BS (1984) Detection of chromogranin in neuroendocrine cells with a monoclonal antibody. *Am J Pathol* 115:458–468
36. Yamada M, Kozuka S, Yamao K, Nakazawa S, Naitoh Y, Tsukamoto Y (1991) Mucin-producing tumor of the pancreas. *Cancer* 68:159–168